

## REVIEW

# The Beneficial Role of Thiamine in Parkinson Disease

Khanh v. q. Lương &amp; Lan T. H. Nguyễn

Vietnamese American Medical Research Foundation, Westminster, CA, USA

**Keywords**

Parkinson disease; Thiamine; Transketolase.

**Correspondence**

K. v. q. Lương, M.D., F.A.C.P., F.A.C.E.,  
F.A.C.N., F.A.S.N., F.C.C.P., and F.A.C.A.A.I.  
(SC), Vietnamese American Medical Research  
Foundation, 14971 Brookhurst Street,  
Westminster, CA 92683, USA.

Tel.: +714-839-5898;

Fax: +714-839-5989;

E-mail: Lng2687765@aol.com

Received 30 November 2012; revision 24

January 2013; accepted 26 January 2013

**SUMMARY**

Parkinson disease (PD) is the second most common form of neurodegeneration among elderly individuals. PD is clinically characterized by tremors, rigidity, slowness of movement, and postural imbalance. In this paper, we review the evidence for an association between PD and thiamine. Interestingly, a significant association has been demonstrated between PD and low levels of serum thiamine, and thiamine supplements appear to have beneficial clinical effects against PD. Multiple studies have evaluated the connection between thiamine and PD pathology, and candidate pathways involve the transcription factor Sp1, p53, Bcl-2, caspase-3, tyrosine hydroxylase, glycogen synthase kinase-3 $\beta$ , vascular endothelial growth factor, advanced glycation end products, nuclear factor kappa B, mitogen-activated protein kinase, and the reduced form of nicotinamide adenine dinucleotide phosphate. Thus, a review of the literature suggests that thiamine plays a role in PD, although further investigation into the effects of thiamine in PD is needed.

doi: 10.1111/cns.12078

**Introduction**

Parkinson disease (PD) is a movement disorder that is characterized by tremor, rigidity, akinesia, and loss of posture reflexes, which often leads to immobility and frequent falls. PD results from the selective loss of dopaminergic (DA) neurons in the substantia nigra (SN) of the brain. Studies have suggested a relationship between dopamine and thiamine, dopamine has been shown to suppress the mouse-killing aggression (muricide) induced by a thiamine-deficient (TD) diet [1], and this suppressive effect can be potentiated by carbidopa [2]. Patients with PD who have undergone levodopa (L-dopa) therapy show significantly higher cerebrospinal fluid (CSF) levels of thiamine diphosphate (TDP) and total thiamine than do patients who are not treated with this drug [3]. Moreover, thiamine deficiency can decrease the concentration of dopamine in the striatum, whereas animals that are fed a diet containing 5% ethanol show increased dopamine turnover [4]. In an animal experimental study of thiamine deficiency, region-specific vesicular dysfunction, that is, a decreased level of dopamine metabolite, was observed posttreatment [5]. Intrastriatal administration of thiamin triphosphate (TTP) or TDP has been shown to induce dopamine release [6], and thiamine derivatives are known to be present in high concentrations in the human SN [7]. Dopamine release can be induced by the intrastriatal administration of TPP or TDP, reaching levels that are as high as 1400% and 249% of the basal levels, respectively, whereas reduced levels of

dopamine in the striatum can occur in thiamine deficiency [6]. Furthermore, decreased CSF-free thiamine levels were noted in patients with PD as compared to controls [3]. In parkinsonism-dementia patients, thiamine pyrophosphatase activity was found to be significantly reduced in the frontal cortex [8]. In addition, Gold et al. [9] reported that 70% and 33% of their patients with PD had low plasma thiamine and low RBC thiamine levels, respectively. Starvation-induced TD encephalopathy may also cause symmetrical lesions in the SN [10]. Together, these findings suggest that thiamine may play a role in DA neuron activity. Interestingly, parental thiamine administration was used successfully in 9 nonalcoholic patients who presented with acute neurological disorders [11]. In addition, the administration of a combination of thiamine and acetazolamide was reported to reduce scores on the Abnormal Involuntary Movement Scale (AIMS) and the Simpson–Angus Neurological Rating Scale (ANRS) in patients with tardive dyskinesia and parkinsonism symptoms [12]. Recently, thiamine has been shown to also improve the symptoms associated with PD; within days of thiamine treatment, patients reportedly had smiles on their faces, walked normally with longer strides, increased their arm swings, and experienced no tremors or sialorrhea. In addition, 3 patients no longer required carbidopa or levodopa and did not suffer ill effects on their movements. [13]. In a previous publication, we identified a number of proteins that link thiamine to PD pathology [14]. In the present paper, we will further discuss the relationship between thiamine and PD.

## The Role of Thiamine in Parkinson Disease

The Sp1 transcription factor is a member of an extended family of DNA-binding proteins that are acetylated in neurons in response to oxidative stress [15]. The Sp1 family of proteins plays an important role in controlling the expression of the dopamine transporter gene within DA neurons [16], and these proteins also regulate the expression of the rat dopamine receptor gene [17]. The rat dopamine receptor contains multiple Sp1-binding sites [17,18]. Sp1 or another protein antigenically related to Sp1 is included in the complex that binds the activator region of the human D<sub>1A</sub> dopamine receptor gene [19]. A novel 130-kDa factor recognizing Sp1-binding sequences in the D<sub>2</sub> gene negative modulator is also found in nuclear extract from the rat striatum [20]. Furthermore, the human monoamine oxidase (MAO) B plays a major role in the degradation of biogenic and dietary amines such as phenylethylamine, benzylamine, dopamine, and tyramine. The human monoamine oxidase B gene was also regulated by Sp1 and Sp3 [21]. Similarly, thiamine uptake in the human intestine occurs via a specialized carrier-mediated mechanism, and the human thiamine transporters (THTRs) are expressed in the intestine and are regulated via *Sp1* promoter elements [22,23]. These findings suggest a link between thiamine, Sp1, and DA transporter and indicate that the *Sp1* family of proteins plays an important role in controlling the expression of the dopamine transporter gene within DA neurons and also regulates the activity of *SLC19A3* gene in transport thiamine.

The p53 gene and protein play critical roles in regulation of the normal cell cycle, cell cycle arrest, and the apoptotic response. p53 is a transcription factor that plays a major role in determining cell fates in response to DNA damage; in the central nervous system (CNS), the function of p53 is to serve as a critical regulator of neuronal cell apoptosis [24]. Specifically, p53 is involved in the dopamine-induced apoptosis of cellular granule neurons [25]. In p53-knockout mice, DA neurons were shown to be more resistant to 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine (MPTP)-induced neurotoxicity than normal neurons [26], and an increase in p53 expression was observed in autopsied tissue from patients with PD [27,28]. Furthermore, levels of p53 immunoreactivity increased following 6-hydroxydopamine (6-OHDA)-induced apoptosis of nigral dopaminergic neurons [29]. The activation of p53 has been reported in animal models of PD, and inhibition of p53 activity was shown to prevent MPTP-induced degeneration of DA neurons [30]. These findings suggest that p53-associated apoptosis may be a common mechanism of cell loss in several important neurodegenerative diseases. In addition, the presence of abundant p53-immunoreactive neurites and glial cell processes appears to be a novel feature of neurodegeneration shared by these distinct diseases. Parkin is a PD-associated gene that contributes to the functions of p53 [31]. Genetic depletion of endogenous parkin increases the expression, activity, and mRNA levels of p53 [32]. Moreover, an increased number of thiamine transporters has been observed in cells that over-express thiamine transport genes (mTHTR-1) and in cells exposed to conditions that induce DNA damage or p53 activation [33]. TDP inhibits p53 binding, whereas thiamine inhibits intracellular p53 activity [34]. In addition, the

expression of p53 is significantly decreased when cultures of retinal neurons from diabetic rats are treated with thiamine [35]. These observations suggest that the transcription factor p53 is activated in PD, which increases the apoptotic response to cellular damage, and that thiamine ameliorates the cellular effects of activated p53.

Bcl-2 is a membrane-bound protein that plays a neuroprotective role in the CNS. Bcl-2 inhibits apoptosis and enhances the survival of newly formed neurons in the normal and ischemic hippocampus [36], and Bcl-2 mRNA and protein expression are developmentally regulated in both the human and murine brains [37,38]. Bcl-2 was shown to inhibit cell death caused by serum and growth-factor withdrawal in a central neural cell line, and it has also been shown to have inhibitory effects on calcium ionophore A23187, glucose withdrawal, membrane peroxidation, and, in some cases, even free-radical-induced damage [39]. Oxidative stress induced by the neurotoxins MPTP, paraquat, maneb, and rotenone causes lipid peroxidation and protein misfolding, which has effects on cell death through members of the Bcl-2 family [40]. MPTP-induced DA neuron toxicity was shown to decrease the expression of Bcl-2 in mouse SN [41], whereas Bcl-2 over-expression was protective against MPTP toxicity [41,42] and 6-OHDA toxicity [43,44]. Interestingly, high cellular concentrations of  $\alpha$ -synuclein have been shown to downregulate Bcl-2 expression [45]. G-protein-coupled receptor kinase 9 (GRK5) has been reported to accumulate in Lewy bodies, which are over-expressed in the  $\alpha$ -synuclein model of PD, and regulate Bcl-2 expression [46]. In addition, glial cell-line-derived neurotrophic factor promoted the survival of grafted midbrain-derived neural stem cells and increases the expression of Bcl-2 in a rat model of PD [47]. The endoplasmic reticulum (ER) chaperone  $\sigma$ -1 receptor (Sig-1R) is cytoprotective against ER stress-induced apoptosis [48], and Sig-1Rs are downregulated in the brains of patients with early stage PD [49]. Dopamine was shown to induce apoptosis in Sig-1R knockdown Chinese Hamster ovary cells, which could be blocked by the over-expression of Bcl-2 [50]. Therefore, decreased Bcl-2 protein levels suggest increased levels of apoptosis in patients with PD. However, pretreatment with B vitamins (B<sub>1</sub>, B<sub>6</sub>, and B<sub>12</sub>) had a protective effect on experimentally induced epilepsy of the mouse brain, which was associated with the induction of Bcl-2 expression within 12 h of treatment [51]. Moreover, thiamine deprivation was shown to increase cell death and reduce Bcl-2 expression during hybridoma cell culture [52]. Benfotiamine improves the functional recovery of infarcted hearts and increases Bcl-2 protein levels [53], and it was also shown to prevent LPS-induced apoptosis and enhance Bcl-2 expression in a mouse macrophage cell line [54]. Furthermore, when human and bovine pericytes were intermittently exposed to high levels of glucose, there was a 50–60% decrease in the Bcl-2-to-Bax expression ratio, and the addition of thiamine and benfotiamine completely reversed this damaging effect [55]. Altogether, these results suggest that thiamine may have a neuroprotective role in PD by increasing expression of the apoptotic inhibitor Bcl-2.

Caspases are cysteinyl aspartate-specific proteases that play a critical role in the regulatory and execution phases of apoptosis [56]. Activation of caspases and the apoptosis of DA neurons have been implicated in the pathogenesis of PD. Activated caspases-3

has been observed in the SN of patients with PD [57,58], and intranigral lipopolysaccharide (LPS) injection was shown to induce the degeneration of DA neurons and increases caspase-3 activation in the rat ventral mesencephalon [59]. Glial-cell-line-derived neurotrophic factors were shown to promote the survival of grafted midbrain-derived neural stem cells, and reduce the expression of caspase-3 in a rat model of PD [47]. Moreover, caspase-3 inhibitors were shown to protect neuronal cells from MPTP-induced apoptosis [60], and gene disruption of caspase-3 prevented MPTP-induced apoptosis in the SN [61]. These findings suggest that caspase-3 activation precedes and is not a consequence of apoptotic cell death in PD. However, thiamine transporter *SLC19A3* gene-transfected breast cancer cells also demonstrated increased levels of apoptosis when exposed to doxorubicin and radiation, and this effect was partially mediated by a caspase-3-dependent pathway [62]. Furthermore, the thiamine deficiency caused by thiamine antagonists was shown to lead to caspase-3-dependent apoptosis in neuronally differentiated rat PC-12 cells [63]. In addition, benfotiamine accelerated the healing of ischemic diabetic limbs in mice via the potentiation of angiogenesis and prevented the induction of pro-apoptotic caspase-3 [64], and this compound was also shown to prevent LPS-induced apoptosis and caspase activation in a mouse macrophage cell line [54]. In addition, sulbutiamine, a highly lipid-soluble synthetic analog of thiamine, was shown to attenuate trophic-factor-deprivation-induced cell death in transformed retinal ganglion cells (RGC-5) and decreases the expression of cleaved caspase-3 [65]. These findings suggest that thiamine may play a role in PD by inhibiting the activity of the apoptotic factor caspase-3.

Tyrosine hydroxylase (TH) is the rate-limiting enzyme in the biosynthesis of dopamine and other catecholamines. In normal human brains, the mRNA levels of human TH<sub>1-2</sub> are much greater than those of human TH<sub>3-4</sub>. Marked and parallel decreases in mRNA levels of human TH<sub>1-4</sub> are found in the SN in PD [66]. Humans and monkeys, having multiple TH isoforms, are more susceptible to MPTP than nonprimate mammals with a single form such as mice and rats, which have low susceptibility to MPTP. Such a difference may suggest the functional significance of TH isoforms in PD. The activity and protein level of TH are decreased to cause DA deficiency in the striatum in PD. However, the homo-specific activity (activity/enzyme protein) of TH is increased. This increase in TH homo-specific activity suggests activation by increased phosphorylation at the N-terminus of the TH protein for a compensatory increase in DA synthesis. This compensatory activation of TH by phosphorylation in the remaining DA neurons may contribute to a further decrease in TH protein and activity in DA neurons in PD, causing a vicious circle of decreasing TH activity, protein level, and DA contents [67]. mRNA level and protein content of TH are markedly decreased in the SN and striatum of the postmortem PD brain [68]. The pathophysiology of PD is largely due to the nigro-striatal DA system, as decreases in TH activity, TH synthesis, and TH mRNA levels in the striatum of patients with PD and PD animal models have been observed [69,70]. During the treatment with chronic low dose MPTP, mice developed dopamine-dependent movement deficits induced by loss of TH-positive nigrostriatal axon terminals [71]. MPTP-treated cats exhibited severe Parkinson-like motor syndrome during the acute period with a marked decrease in TH-immunoreactivity in the

striatum [72]. The gait variability in the PD mice showed a closer correlation with the protein levels of TH in the SN than the walking distances in the conventional open field test [73]. The L-dopa-induced increase in striatal TH-immunoreactive neurons is dose dependent and persists for days after L-dopa withdrawal [74]. In addition, TH gene mutations have been reported to be associated with PD; a TH heterozygous variant was reported in one patient with dopa-responsive dystonia simulating spastic paraplegia [75] as well as in early-onset patients with PD [76]. In addition, a novel deletion in the TH gene was detected in one patients with PD [77], and decreased levels of TH protein were noted in the striatum of the MPTP-induced neurotoxic lesions of animals with experimental PD [78]. Moreover, the fluorescence intensity of TH expression was decreased in the limbic cortex and brainstem in TD mice compared with pair-fed mice as the control group [79]. In addition, male Wistar rats maintained on a TD diet demonstrated mouse-killing behavior, which was attenuated by the administration of L-dopa [80]. Similarly, this suppressive effect was shown to be potentiated by carbidopa [2].

Glycogen synthase kinase-3 $\beta$  (GSK3 $\beta$ ) is a protein kinase that is involved in many physiological processes (e.g., metabolism, gene expression, and apoptosis). GSK3 $\beta$  is pivotal in controlling neuronal polarity within primary embryonic hippocampal neurons [81]. GSK3 $\beta$  is associated with the fate of DA neurons in PD and may exert its toxicity via the induction of apoptosis by the direct activation of intrinsic cascades or via the phosphorylation of synphilin-1 or  $\alpha$ -synuclein. GSK3 $\beta$  expression is increased in brain regions associated with PD pathology [82]. In animal and cell culture models of PD, rotenone-induced cytotoxicity is mediated by microtubule destabilization via GSK3 $\beta$  activation, and that microtubule destabilization is caused by reduction in the binding capacity of tau to microtubules, which is a result of tau phosphorylation via GSK3 $\beta$  activation. Rotenone-induced cytotoxicity in SH-SY5Y cells was attenuated by the GSK3 $\beta$  inhibitor SB216763 [83]. GSK3 $\beta$  polymorphisms alter transcription and splicing by interacting with Tau haplotypes to modify disease risk in patients with PD [84]. Haplotype analysis revealed that the TT haplotype of GSK3 $\beta$  polymorphisms was over-represented in patients with PD as compared to controls [85]. However, GSK3 $\beta$  variant reduces the risk of PD in Han Chinese population [86]. The GSK3 inhibitor reduces L-dopa-induced neurotoxicity [87], and exposure to pyriethamine, an antithiamine compound, increases A $\beta$  accumulation and GSK3 activity in the brain [88]. In an animal model of AD, benfotiamine was shown to improve cognitive function, reduce amyloid deposition, and suppress GSK3 activity [89]. These findings suggest that thiamine may play a role in PD by suppressing GSK3 activity.

Angiogenesis is a complex process that involves coordinated endothelial cell activation, proliferation, migration, tube formation, and capillary sprouting. In addition, angiogenesis requires the participation of numerous intracellular signaling pathways. Vascular endothelial growth factor (VEGF) is a key mediator of angiogenesis and has been shown to have neuroprotective effects on DA neurons in models of 6-OHDA-induced toxicity, to decrease amphetamine-induced rotational behavior, and to preserve TH-positive neurons and fibers [90]. In rat midbrain cultures, increased levels of VEGF-B transcription have been reported following the addition of the neurotoxin rotenone [91], suggesting that the growth factor VEGF-B can improve neuronal survival in a

culture model of PD. An increase in the number of VEGF-positive neurons and blood vessels was also been demonstrated in the SN of mice with MPTP-induced neurotoxicity [92]. These changes in vascularization may therefore modify the neuronal availability of blood nutrients, blood cells or toxic substances, and neuronal susceptibility to parkinsonism. Wada et al. [93] further demonstrated upregulated expression of VEGF in the SN of patients with PD. In PD animal models, the neuroprotective effects of VEGF appear to be dose dependent. Indeed, low doses of VEGF have been shown to have a neuroprotective effect on DA neurons and have been shown to result in behavioral improvement, whereas high doses have been shown to induce angiogenesis and glial proliferation [94]. Moreover, thiamine deficiency was shown to result in polyneuropathy after gastrectomy, and this deficiency has also been associated with high levels of serum VEGF, which typically returned to normal following the intravenous administration of thiamine, which further improves symptoms of polyneuropathy [95]. In addition, increased serum levels of VEGF have been reported in patients with wet beriberi [96]. In a model of peritoneal dialysis in uremic rats, treatment with benfotiamine decreased peritoneal fibrosis, markers of inflammation, neovascularization, and VEGF staining [97]. Furthermore, benfotiamine was also shown to improve the functional recovery of infarcted hearts and also reduce the phosphorylation/activation of VEGF receptor 2/Akt signaling pathways in a mouse macrophage cell line [53].

Glyoxalase 1 (Glyo-1) catalyzes the initial rate-limiting step in the removal of methylglyoxal (MG), which is the major precursor for advanced glycation end products (AGEs). AGEs represent a heterogeneous group of macromolecules that are formed by the nonenzymatic glycation of proteins, lipids, and nucleic acids. RAGEs are multiligand receptors, and their ligands also likely recognize several receptors to mediate their numerous biological effects [98].  $\alpha$ -Synuclein has been implicated in PD, and its deficiency leads to increased Glyo-1 expression and glycation stress [99]. Altered Glyo-1 expression was also reported in brain of mouse with parkin deficiency [100]. Furthermore, glycation was observed in the SN and locus ceruleus, with the greatest levels of immunoreactivity at the periphery of Lewy bodies, in patients with PD [101]. In addition, AGEs were shown to stimulate the *in vitro* cross-linking of  $\alpha$ -synuclein and accelerate intracellular inclusion body formation [102], and RAGE levels were found to be over-expressed in patients with PD as compared to age-matched controls [103]. RAGE deficiency protects nigral DA neurons against cell death induced by the neurotoxin MPTP, and this type of cell death mimics many of the characteristic features of PD [104]. Moreover, thiamine and benfotiamine supplementation prevented the tissue accumulation and increased the urinary excretion of protein glycation, oxidation, and nitration adducts associated with experimental diabetes [105]. Karachalias et al. [106] reported that the hydroimidazolone of AGE residues derived from glyoxal and methylglyoxal (G-H1 and MG-H1) was increased by 115% and 68%, respectively, in streptozotocin-induced diabetic rats, whereas treatment with thiamine and benfotiamine normalized these effects. However, N-carboxymethyl-lysine (CML) and N-carboxyethyl-lysine (CEL) residues increased by 74% and 118%, respectively, in diabetic-induced rats, and only treatment with thiamine normalized these effects. In addition,

serum markers of endothelial dysfunction, oxidative stress, and AGEs were shown to be increased following a meal high in AGE content, although benfotiamine significantly reduced these effects [107]. The addition of benfotiamine was also shown to enhance transketolase activity and decrease the expression of AGEs and RAGEs in a model of peritoneal dialysis in uremic rats [97]. In both bovine aortic endothelial cells and the retinas of diabetic rats, benfotiamine inhibited the AGE formation pathway by activating transketolase and prevented experimental diabetic retinopathy [108]. Furthermore, the combined administration of thiamine and vitamin B6 to patients with diabetic nephropathy decreased DNA glycation in leukocytes, although vitamin B6 alone did not have such an effect [109].

The transcription factor nuclear factor kappa B (NF- $\kappa$ B) is a hetero-dimeric, sequence-specific transcription factor that is found in many cell types. NF- $\kappa$ B has been implicated in chronic inflammatory diseases, and it is a key regulator of genes involved in responses to infection, inflammation, and stress. Increased activation of NF- $\kappa$ B has been reported in DA neurons of the SN in patients with PD as compared to controls [28,110,111]. NF- $\kappa$ B has also been identified as a component of Lewy bodies [112]. A significant increase in NF- $\kappa$ B was observed mainly in glial cells of the SN during MPTP-induced apoptosis in a mouse model of PD [113]. Furthermore,  $\alpha$ -synuclein over-expression was shown to enhance manganese-induced neurotoxicity via the NF- $\kappa$ B-mediated pathway [114]. Selective inhibition of NF- $\kappa$ B activation was shown to suppress nigral microglial activation and improve motor function in a mouse model of PD [115], and benfotiamine was also shown to inhibit NF- $\kappa$ B activation, via the activation of transketolase, and prevent experimental diabetic retinopathy in both bovine aortic endothelial cells and the retinas of diabetic rats [108]. Benfotiamine was further shown to prevent endotoxin-induced inflammation by suppressing oxidative-stress-induced NF- $\kappa$ B activation in rats with endotoxin-induced uveitis and in murine macrophage cell lines [54,116], and benfotiamine-mediated suppression of expression of NF- $\kappa$ B prevented LPS-induced macrophage cell death and monocyte adhesion to endothelial cells [117]. Altogether, these findings indicate that thiamine may suppress NF- $\kappa$ B activation in PD.

The mitogen-activated protein kinase (MAPK) pathways provide a key link between membrane-bound receptors and changes in gene expression, involving the extracellular signal-regulated kinase (ERK) cascade, the stress-activated protein kinases/c-jun N-terminal kinase (SAPK/JNK) cascade, and the p38 MAPK/RK/HOG cascade [118]. Increased cytoplasmic ERK1/2 activity has been observed in the brains of human patients with PD [119], and degenerating SN neurons typically display phosphorylated-ERK1/2 granules [120]. The activation of ERK1/2 is induced by the neurotoxin 6-OHDA, and inhibition of ERK activation enhances neuronal survival [121,122]. The mitochondrial localization of ERK2 activity suggests an effect of 6-OHDA on mitophagy and autophagic cell death in PD [123]. Dysregulation of the autophagy pathway has been observed in the brains of patients with PD and in animal models of PD [124,125]. In addition, the activation of p38 MAPK has been demonstrated in the SN of MPTP-treated mouse models of PD [126]. Moreover, vulnerability to glutamate-induced toxicity in DA neurons is dependent on endogenous dopamine



as well as MAPK activation [126]. Interestingly, genetic deficiency in MAPK kinase  $2^{-/-}$  prevented MPTP-induced neurotoxicity in mouse models of PD [127]. Moreover, benfotiamine was shown to modulate the macrophage response to bacterial endotoxin-induced inflammation by preventing the activation of p-38 MAPK and stress-activated kinases (SAPK/JNK) [54].

The reduced form of the nicotinamide adenine dinucleotide phosphate (NADPH) oxidase (NOX) enzyme complex mediates critical physiological and pathological processes including cell signaling, inflammation, and mitogenesis, by generating reactive oxygen species (ROS) from molecular oxygen. NOX is widely expressed in various immune cells, including microglia, macrophages, and neutrophils. NOX expression was observed in the nuclei of DA neurons in the SN of patients with PD and animals with 6-OHDA-induced neurotoxicity [128]. NOX activation was reported to increase zinc-induced DA neurodegeneration as well as MPPT, rotenone, angiotensin, and paraquat-induced neurotoxicity in animal models of PD [129–133]. Inhibition and knock-down of NOX were shown to reduce paraquat-induced ROS generation and DA cell death [133], and NOX inhibitors were also shown to protect against LPS-induced toxicity, MPPT-induced oxidative stress, and apoptosis in mesencephalic DA neuronal cells [134,135]. Thiamine is an essential coenzyme for transketolase, which is part of the pentose phosphate pathway that helps maintain cellular NADPH levels. In a study that administered glyoxal toxicity to hepatocytes, thiamin demonstrated cytoprotective functions and restored NADPH levels, glyoxal detoxification, and mitochondrial membrane potential [136]. Furthermore, NADPH-cytochrome c reductase levels were increased in TD animals [137], and benfotiamine treatment under both normo- and hyper-glycemic conditions significantly downregulated Nox4 expression [138]. In addition, animals that were fed a high-thiamine diet had approximately 57% of the NADPH-cytochrome c reductase activity of those that were fed a TD diet [139]. Alto-

gether, these results suggest that thiamine may be neuroprotective against PD by regulating NADPH-cytochrome c activity.

## Conclusions

Thiamine plays a beneficial role in PD by inducing dopamine release and improving the symptoms associated with PD. Genetic studies have provided the opportunity to identify the specific proteins that link thiamine to the pathology of PD. Thiamine also exerts its effects on PD via nongenomic mechanisms. In addition, thiamine involved in PD, including the DJ-1 gene, excitatory amino acid transporters (EAATs), the  $\alpha$ -ketoglutarate dehydrogenase complex, coenzyme Q10, lipoamide dehydrogenase, chromosome 7, transcription factor p53, the renin-angiotensin system, heme oxygenase-1, and *poly(ADP-ribose) polymerase-1* gene [14]. However, gastrointestinal dysfunction is common in patients with PD, and it potentially affects the therapeutic intervention [140]. Gastric emptying has been reported to be frequently delayed in patients with PD [141]. Decreased nonmediated uptake across the enterocyte brush border membrane was demonstrated in patients with PD [142]. In addition, the intestinal absorption of thiamine is sufficient in young people but may be reduced with age [143]. Parental administration of thiamine may be suitable for patients with PD [14]. Thus, further studies are needed to determine the potential benefits of using thiamine as a treatment for PD.

## Funding

The authors, Dr. Khanh Luong and Dr. Lan Nguyen, received no funding for this study.

## Conflict of Interest

The authors declare no conflict of interest.

## References

- Tadano T, Abe Y, Morikawa Y, et al. Involvement of dopaminergic neurons in mouse-killing aggression in rats. *Methods Find Exp Clin Pharmacol* 1997;**19**:527–531.
- Onodera K. Effects of decarboxylase inhibitors on muricidal suppression by L-dopa in thiamine deficient rats. *Arch Int Pharmacodyn Ther* 1987;**285**:263–276.
- Jiménez-Jiménez FJ, Molina JA, Hernández A, et al. Cerebrospinal fluid levels of thiamine in patients with Parkinson's disease. *Neurosci Lett* 1999;**271**:33–36.
- Sjöquist B, Johnson HA, Neri A, Lindén S. The influence of thiamine deficiency and ethanol on rat brain catecholamines. *Drug Alcohol Depend* 1988;**22**:167–193.
- Mousseau DD, Raghavendra Rao VL, Butterworth RF. Vesicular dysfunction during experimental thiamine deficiency is indicated by alterations in dopamine metabolism. *Eur J Pharmacol* 1996;**317**:263–267.
- Yamashita H, Zhang Y-X, Nakamura S. The effects of thiamine and its phosphate esters on dopamine release in the rat striatum. *Neurosci Lett* 1993;**158**:229–231.
- Baker H, Frank O, Chen T, Feingold S, DeAngelis B, Baker E. Vitamin content of some normal human brain segments. *J Neurosci Res* 1984;**11**:419–435.
- Laforenza U, Patrini C, Poloni M, et al. Thiamine-mono- and pyrophosphatase activities from brain homogenate of Guamanian amyotrophic lateral sclerosis and parkinsonism-dementia patients. *J Neurol Sci* 1992;**109**:156–161.
- Gold M, Hauser RA, Chen MF. Plasma thiamine deficiency associated with Alzheimer's disease but not Parkinson's disease. *Metab Brain Dis* 1998;**13**:43–53.
- Kalidass B, Sunnathkal R, Rangashamanna DV, Paraswani R. Atypical Wernicke's encephalopathy showing involvement of substantia nigra. *J Neuroimaging* 2012;**22**:204–207.
- Merkin-Zaborsky H, Ifergane G, Frisher S, Valdman S, Herishanu Y, Wirguin I. Thiamine-responsive acute neurological disorders in nonalcoholic patients. *Eur Neurol* 2001;**45**:34–37.
- Cowen MA, Green M, Bertollo DN, Abbott K. A treatment for tardive dyskinesia and some other extrapyramidal symptoms. *J Clin Psychopharmacol* 1997;**17**:190–193.
- Lương K, Nguyễn L. The beneficial role of Thiamine in Parkinson's disease: preliminary report. *J Neurol Res* 2012;**2**:211–214.
- Luong Kv, Nguyễn LT. Thiamine and Parkinson's disease. *J Neurol Sci* 2012;**316**:1–8.
- Ryu H, Lee J, Zaman K, et al. Sp1 and Sp3 are oxidative stress-inducible, antideath transcription factors in cortical neurons. *J Neurosci* 2003;**23**:3597–3606.
- Wang J, Bannon MJ. Sp1 and Sp3 activate transcription of the human dopamine transporter gene. *J Neurochem* 2005;**93**:474–482.
- Yajima S, Lee SH, Minowa T, Mouradian MM. Sp family transcription factors regulate expression of rat D2 dopamine receptor gene. *DNA Cell Biol* 1998;**17**:471–479.
- Healy DP, O'Rourke DA. Regulation of dopamine-1A ( $D_{1A}$ ) receptor gene transcription. *Clin Exp Hypertens* 1997;**19**:1–13.
- Minowa MT, Minowa T, Mouradian MM. Activator region analysis of the human  $D_{1A}$  dopamine receptor gene. *J Biol Chem* 1993;**268**:23544–23551.
- Minowa T, Minowa MT, Mouradian MM. Negative modulator of the rat  $D_2$  dopamine receptor gene. *J Biol Chem* 1994;**269**:11656–11662.
- Wong WK, Chen K, Shih JC. Regulation of human monoamine oxidase B gene by Sp1 and Sp3. *Mol Pharmacol* 2001;**59**:852–859.
- Nabokina SM, Said HM. Characterization of the 5'-regulatory region of the human thiamin transporter SLC19A3: *in vitro* and *in vivo* studies. *Am J Physiol Gastrointest Liver Physiol* 2004;**287**:G822–G829.
- Nabokina SM, Reidling JC, Said HM. Differentiation-dependent up-regulation of intestinal thiamin uptake: cellular and molecular mechanisms. *J Biol Chem* 2005;**280**:32676–32682.
- Araki N, Morimasa T, Sakai T, et al. Comparative analysis of brain proteins from p53-deficient mice by two-dimensional electrophoresis. *Electrophoresis* 2002;**21**:1880–1889.

25. Daily D, Barzilai A, Offen D, Kamsler A, Melamed E, Ziv I. The involvement of p53 in dopamine-induced apoptosis of cerebellar granule neurons and leukemic cells overexpressing p53. *Cell Mol Neurobiol* 1999;**19**:261–276.
26. Trimmer PA, Smith TS, Jung AB, Bennett JP Jr. Dopamine neurons from transgenic mice with a knockout of the p53 gene resist MPTP neurotoxicity. *Neurodegeneration* 1996;**5**:233–239.
27. de la Monte SM, Sohn YK, Ganju N, Wands JR. P53- and CD95-associated apoptosis in neurodegenerative diseases. *Lab Invest* 1998;**78**:401–411.
28. Mogi M, Kondo T, Mizuno Y, Nagatsu T. p53 protein, interferon-gamma, and NF-kappaB levels are elevated in the parkinsonian brain. *Neurosci Lett* 2007;**414**:94–97.
29. Liang ZQ, Li YL, Zhao XL, et al. NF-kappaB contributes to 6-hydroxydopamine-induced apoptosis of nigral dopaminergic neurons through p53. *Brain Res* 2007;**1145**:190–203.
30. Biswas SC, Ryu E, Park C, Malagelada C, Greene LA. Puma and p53 play required roles in death evoked in a cellular model of Parkinson disease. *Neurochem Res* 2005;**30**:839–845.
31. Zhang C, Lin M, Wu R, et al. Parkin, a p53 target gene, mediates the role of p53 in glucose metabolism and the Warburg effect. *Proc Natl Acad Sci USA* 2011;**108**:16259–16264.
32. da Costa CA, Sunyach C, Giaime E, et al. Transcriptional repression of p53 by parkin and impairment by mutations associated with autosomal recessive juvenile Parkinson's disease. *Nat Cell Biol* 2009;**11**:1370–1375.
33. Lo PK, Chen JY, Tang PP, et al. Identification of a mouse thiamine transporter gene as a direct transcriptional target for p53. *J Biol Chem* 2001;**276**:37186–37193.
34. McLure KG, Takagi M, Kastan MB. NAD<sup>+</sup> modulates p53 DNA binding specificity and function. *Mol Cell Biol* 2004;**24**:9958–9967.
35. Yang Z, Ge J, Yin W, Shen H, Liu H, Guo Y. The expression of p53, MDM2 and Ref1 gene in cultured retina neurons of SD rats treated with vitamin B1 and/or elevated pressure. *Yan Ke Xue Bao* 2004;**20**:259–263. [Article in Chinese]
36. Sasaki T, Kitagawa K, Yagita Y, et al. Bcl2 enhances survival of newborn neurons in the normal and ischemic hippocampus. *J Neurosci Res* 2006;**84**:1187–1196.
37. Jarskog LF, Gilmore JH. Developmental expression of Bcl-2 protein in human cortex. *Brain Res Dev Brain Res* 2000;**119**:225–230.
38. Shimohama S, Fujimoto S, Sumida Y, Tanino H. Differential expression of rat brain bcl-2 family proteins in development and aging. *Biochem Biophys Res Commun* 1998;**252**:92–96.
39. Zhong LT, Sarafian T, Kane DJ, et al. bcl-2 inhibits death of central neural cells induced by multiple agents. *Proc Natl Acad Sci USA* 1993;**90**:4533–4537.
40. Ethell DW, Fei Q. Parkinson-linked genes and toxins that affect neuronal cell death through the Bcl-2 family. *Antioxid Redox Signal* 2009;**11**:529–540.
41. Vila M, Jackson-Lewis V, Vukosavic S, et al. Bax ablation prevents dopaminergic neurodegeneration in the 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine mouse model of Parkinson's disease. *Proc Natl Acad Sci USA* 2001;**98**:2837–2842.
42. Offen D, Beart PM, Cheung NS, et al. Transgenic mice expressing human Bcl-2 in their neurons are resistant to 6-hydroxydopamine and 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine neurotoxicity. *Proc Natl Acad Sci USA* 1998;**95**:5789–5794.
43. Yamada M, Oligino T, Mata M, Goss JR, Glorioso JC, Fink DJ. Herpes simplex virus vector-mediated expression of Bcl-2 prevents 6-hydroxydopamine-induced degeneration of neurons in the substantia nigra in vivo. *Proc Natl Acad Sci USA* 1999;**96**:4078–4083.
44. Natsume A, Mata M, Goss J, et al. Bcl-2 and GDNF delivered by HSV-mediated gene transfer act additively to protect dopaminergic neurons from 6-OHDA-induced degeneration. *J Neurotrauma* 2002;**19**:61–68.
45. Seo JH, Rah JC, Choi SH, et al. Alpha-synuclein regulates neuronal survival via Bcl-2 family expression and PI3/Akt kinase pathway. *FASEB J* 2002;**16**:1826–1828.
46. Liu P, Wang X, Gao N, et al. G protein-coupled receptor kinase 5, overexpressed in the alpha-synuclein up-regulation model of Parkinson's disease, regulates bcl-2 expression. *Brain Res* 2010;**1307**:134–141.
47. Lei Z, Jiang Y, Li T, Zhu J, Zeng S. Signaling of glial cell line-derived neurotrophic factor and its receptor GFR $\alpha$ 1 induce Nurr1 and Ptx3 to promote survival of grafted midbrain-derived neural stem cells in a rat model of Parkinson disease. *J Neuropathol Exp Neurol* 2011;**70**:736–747.
48. Hayashi T, Justinova Z, Hayashi E, et al. Regulation of sigma-1 receptors and endoplasmic reticulum chaperones in the brain of methamphetamine self-administering rats. *J Pharmacol Exp Ther* 2010;**332**:1054–1063.
49. Mishina M, Ishiwata K, Ishii K, et al. Function of signal receptors in Parkinson's disease. *Acta Neurol Scand* 2005;**112**:103–107.
50. Mori T, Hayashi T, Su TP. Compromising  $\sigma$ -1 receptors at the endoplasmic reticulum render cytotoxicity to physiologically relevant concentrations of dopamine in a nuclear factor- $\kappa$ B/Bcl-2-dependent mechanism: potential relevance to Parkinson's disease. *J Pharmacol Exp Ther* 2012;**341**:663–671.
51. Rabie T, Mülhofer W, Bruckner T, et al. Transient protective effect of B-vitamins in experimental epilepsy in the mouse brain. *J Mol Neurosci* 2010;**41**:74–79.
52. Ishaque A, Al-Rubeai M. Role of vitamins in determining apoptosis and extent of suppression by bcl-2 during hybridoma cell culture. *Apoptosis* 2002;**7**:231–239.
53. Katare R, Caporali A, Emanuelli C, Madeddu P. Benfotiamine improves functional recovery of the infarcted heart via activation of pro-survival G6PD/Akt signaling pathway and modulation of neurohormonal response. *J Mol Cell Cardiol* 2010;**49**:625–638.
54. Yadav UC, Kalariya NM, Srivastava SK, Ramana KV. Protective role of benfotiamine, a fat-soluble vitamin B1 analogue, in lipopolysaccharide-induced cytotoxic signals in murine macrophages. *Free Radic Biol Med* 2010;**48**:1423–1434.
55. Beltramo E, Berrone E, Tarallo S, Porta M. Different apoptotic responses of human and bovine pericytes to fluctuating glucose levels and protective role of thiamine. *Diabetes Metab Res Rev* 2009;**25**:566–576.
56. Salvesen GS, Riedl SJ. Caspase mechanisms. *Adv Exp Med Biol* 2008;**615**:13–23.
57. Hartmann A, Hunot S, Michel PP, et al. Caspase-3: a vulnerability factor and final effector in apoptotic death of dopaminergic neurons in Parkinson's disease. *Proc Natl Acad Sci USA* 2000;**97**:2875–2880.
58. Tatton NA. Increased caspase 3 and Bax immunoreactivity accompany nuclear GAPDH translocation and neuronal apoptosis in Parkinson's disease. *Exp Neurol* 2000;**166**:29–43.
59. Burguillos MA, Hajji N, Englund E, et al. Apoptosis-inducing factor mediates dopaminergic cell death in response to LPS-induced inflammatory stimulus: evidence in Parkinson's disease patients. *Neurobiol Dis* 2011;**41**:177–188.
60. Dodel RC, Du Y, Bales KR, Ling ZD, Carvey PM, Paul SM. Peptide inhibitors of caspase-3-like proteases attenuate 1-methyl-4-phenylpyridinium-induced toxicity of cultured fetal rat mesencephalic dopamine neurons. *Neuroscience* 1998;**86**:701–707.
61. Yamada M, Kida K, Amutuhair W, Ichinose F, Kaneki M. Gene disruption of caspase-3 prevents MPTP-induced Parkinson's disease in mice. *Biochem Biophys Res Commun* 2010;**402**:312–318.
62. Liu S, Huang H, Lu X, et al. Down-regulation of thiamine transporter THTR2 gene expression in breast cancer and its association with resistance to apoptosis. *Mol Cancer Res* 2003;**1**:665–673.
63. Chorny S, Parkhomenko J, Chorna N. Thiamine deficiency caused by thiamine antagonists triggers upregulation of apoptosis inducing factor gene expression and leads to caspase 3-mediated apoptosis in neuronally differentiated rat PC-12 cells. *Acta Biochim Pol* 2007;**54**:315–322.
64. Gadai S, Emanuelli C, Van Linthout S, et al. Benfotiamine accelerates the healing of ischaemic diabetic limbs in mice through protein kinase B/Akt-mediated potentiation of angiogenesis and inhibition of apoptosis. *Diabetologia* 2006;**49**:405–420.
65. Kang KD, Majid AS, Kim KA, et al. Sulbutiamine counteracts trophic factor deprivation induced apoptotic cell death in transformed retinal ganglion cells. *Neurochem Res* 2010;**35**:1828–1839.
66. Ichinos H, Ohly T, Fujita K, et al. Quantification of mRNA of tyrosine hydroxylase and aromatic L-amino acid decarboxylase in the substantia nigra in Parkinson's disease and schizophrenia. *J Neural Transm* 1994;**8**:149–158.
67. Nakashima A, Ota A, Kaneko YS, Mori K, Nagasaki H, Nagatsu T. A possible pathophysiological role of tyrosine hydroxylase in Parkinson's disease suggested by postmortem brain biochemistry: a contribution for the special 70th birthday symposium in honor of Prof. Peter Riederer. *J Neural Transm* 2013;**120**:49–54.
68. Nagatsu T, Sawada M. Biochemistry of postmortem brains in Parkinson's disease: historical overview and future prospects. *J Neural Transm Suppl* 2007;**113**:120.
69. Zhu Y, Zhang J, Zeng Y. Overview of tyrosine hydroxylase in Parkinson's disease. *CNS Neurol Disord Drug Targets* 2012;**11**:350–358.
70. Feve AP. Current status of tyrosine hydroxylase in management of Parkinson's disease. *CNS Neurol Disord Drug Targets* 2012;**11**:450–455.
71. Korecka JA, Eggers R, Swaab DF, Bossers K, Verhaagen J. Modeling early Parkinson's disease pathology with chronic low dose MPTP treatment. *Restor Neurol Neurosci* 2013;**31**:155–167.
72. Aznavour N, Cendres-Bozzi C, Lemoine L, et al. MPTP animal model of Parkinsonism: dopamine cell death or only tyrosine hydroxylase impairment? A study using PET imaging, autoradiography, and immunohistochemistry in the cat. *CNS Neurosci Ther* 2012;**18**:934–941.
73. Wang XH, Lu G, Hu X, et al. Quantitative assessment of gait and neurochemical correlation in a classical murine model of Parkinson's disease. *BMC Neurosci* 2012;**13**:142.
74. Espadas I, Darmopil S, Vergara-Vera E, et al. L-DOPA-induced increase in TH-immunoreactive striatal neurons in parkinsonian mice: insights into regulation and function. *Neurobiol Dis* 2012;**48**:271–281.
75. Furukawa Y, Graf WD, Wong H, Shimadzu M, Kish SJ. Dopa-responsive dystonia simulating spastic paraplegia due to tyrosine hydroxylase (TH) gene mutations. *Neurology* 2001;**56**:260–263.
76. Hertz JM, Ostergaard K, Juncker I, et al. Low frequency of Parkin, tyrosine hydroxylase, and GTP Cyclohydrolase I gene mutations in a Danish population of early-onset Parkinson's disease. *Eur J Neurol* 2006;**13**:385–390.
77. Bademci G, Edwards TL, Torres AL, et al. A rare novel deletion of the tyrosine hydroxylase gene in Parkinson disease. *Hum Mutat* 2010;**31**:E1767–E1771.
78. Domenger D, Dea D, Theroux L, Moquin L, Gratton A, Poirier J. The MPTP neurotoxic lesion model of Parkinson's disease activates the apolipoprotein E

- cascade in the mouse brain. *Exp Neurol* 2012;**233**:513–522.
79. Nakagawasai O, Yamadera F, Iwasaki K, et al. Preventive effect of kami-untan-to on performance in the forced swimming test in thiamine-deficient mice: relationship to functions of catecholaminergic neurons. *Behav Brain Res* 2007;**177**:315–321.
  80. Abe Y, Tadano T, Yonezawa A, Kisara K. Suppressive effects of intraventricular injected dopamine and nomifensine on muricide induced by thiamine deficiency. *Pharmacol Biochem Behav* 1987;**26**:77–81.
  81. Jiang H, Guo W, Liang X, Rao Y. Both the establishment and the maintenance of neuronal polarity require active mechanisms: critical roles of GSK-3 $\beta$  and its upstream regulators. *Cell* 2005;**120**:123–135.
  82. Nagao M, Hayashi H. Glycogen synthase kinase-3 $\beta$  is associated with Parkinson's disease. *Neurosci Lett* 2009;**449**:103–107.
  83. Hongo H, Kihara T, Kume T, et al. Glycogen synthase kinase-3 $\beta$  activation mediates rotenone-induced cytotoxicity with the involvement of microtubule destabilization. *Biochem Biophys Res Commun* 2012;**426**:94–99.
  84. Kwok JB, Hallupp M, Loy CT, et al. GSK3B polymorphisms alter transcription and splicing in Parkinson's disease. *Ann Neurol* 2005;**58**:829–839.
  85. Kalinderi K, Fidan L, Katsarou Z, Clarimón J, Bostantjopoulou S, Kotsis A. GSK3 $\beta$  polymorphisms, MAPT H1 haplotype and Parkinson's disease in a Greek cohort. *Neurobiol Aging* 2011;**32**:546.
  86. Zhao DM, Li NN, Zhang JH, et al. GSK3 $\beta$  reduces risk of sporadic Parkinson's disease in ethnic Chinese. *Am J Med Genet B Neuropsychiatr Genet* 2012;**159B**:718–721.
  87. Koh SH, Song C, Noh MY, et al. Inhibition of glycogen synthase kinase-3 reduces L-DOPA-induced neurotoxicity. *Toxicology* 2008;**247**:112–118.
  88. Zhao J, Sun X, Yu Z, et al. Exposure to pyrimidine increases beta-amyloid accumulation, Tau hyperphosphorylation, and glycogen synthetase kinase-3 activity in the brain. *Neurotox Res* 2010;**19**:575–583.
  89. Pan X, Gong N, Zhao J, et al. Powerful beneficial effects of benfotiamine on cognitive impairment and beta-amyloid deposition in amyloid precursor protein/presenilin-1 transgenic mice. *Brain* 2010;**133**:1342–1351.
  90. Yasuhara T, Shingo T, Kobayashi K, et al. Neuroprotective effects of vascular endothelial growth factor (VEGF) upon dopaminergic neurons in a rat model of Parkinson's disease. *Eur J Neurosci* 2004;**19**:1494–1504.
  91. Falk T, Zhang S, Sherman SJ. Vascular endothelial growth factor B (VEGF-B) is up-regulated and exogenous VEGF-B is neuroprotective in a culture model of Parkinson's disease. *Mol Neurodegener* 2009;**4**:49.
  92. Barcia C, Bautista V, Sánchez-Bahillo A, et al. Changes in vascularization in substantia nigra pars compacta of monkeys rendered parkinsonian. *J Neural Transm* 2005;**112**:1237–1248.
  93. Wada K, Arai H, Takamashi M, et al. Expression levels of vascular endothelial growth factor and its receptors in Parkinson's disease. *NeuroReport* 2006;**17**:705–709.
  94. Yasuhara T, Shingo T, Muraoka K, et al. The differences between high and low-dose administration of VEGF to dopaminergic neurons of *in vitro* and *in vivo* Parkinson's disease model. *Brain Res* 2005;**1038**:1–10.
  95. Nakagawa H, Yoneda M, Maeda A, Umehara F, Kuriyama M. [Thiamine deficiency polyneuropathy after gastrectomy associated with high level of serum vascular endothelial growth factor (VEGF). A case report]. *Rinsho Shinkeigaku* 2004;**44**:91–95. Article in Japanese
  96. Imai N, Kubota M, Saitou M, Yagi N, Serizawa M, Kobari M. Increase of serum vascular endothelial growth factors in wet beriberi: two case reports. *Intern Med* 2012;**51**:929–932.
  97. Kihm LP, Müller-Krebs S, Klein J, et al. Benfotiamine protects against peritoneal and kidney damage in peritoneal dialysis. *J Am Soc Nephrol* 2011;**22**:914–926.
  98. Bierhaus A, Humpert PM, Morcos M, et al. Understanding RAGE, the receptor for advanced glycation end products. *J Mol Med (Berl)* 2005;**83**:876–886.
  99. Kurz A, Rabbani N, Walter M, et al. Alpha-synuclein deficiency leads to increased glyoxalase I expression and glycation stress. *Cell Mol Life Sci* 2011;**68**:721–733.
  100. Palacino JJ, Sagi D, Goldberg MS, et al. Mitochondrial dysfunction and oxidative damage in parkin-deficient mice. *J Biol Chem* 2004;**279**:18614–18622.
  101. Castellani R, Smith MA, Richey PL, Perry G. Glycoxidation and oxidative stress in Parkinson disease and diffuse Lewy body disease. *Brain Res* 1996;**737**:195–200.
  102. Shaikh S, Nicholson LF. Advanced glycation end products induce *in vitro* cross-linking of alpha-synuclein and accelerate the process of intracellular inclusion body formation. *J Neurosci Res* 2008;**86**:2071–2082.
  103. Dalfó E, Portero-Otín M, Ayala V, Martínez A, Pamplona R, Ferrer I. Evidence of oxidative stress in the neocortex in incidental Lewy body disease. *J Neuropathol Exp Neurol* 2005;**64**:816–830.
  104. Teismann P, Sathe K, Bierhaus A, et al. Receptor for advanced glycation endproducts (RAGE) deficiency protects against MPTP toxicity. *Neurobiol Aging* 2012;**33**:2478–2490.
  105. Karachalias N, Babaei-Jadidi R, Rabbani N, Thornalley PJ. Increased protein damage in renal glomeruli, retina, nerve, plasma and urine and its prevention by thiamine and benfotiamine therapy in a rat model of diabetes. *Diabetologia* 2010;**53**:1506–1516.
  106. Karachalias N, Babaei-Jadidi R, Kupich C, Ahmed N, Thornalley PJ. High-dose thiamine therapy counters dyslipidemia and advanced glycation of plasma protein in streptozotocin-induced diabetic rats. *Ann N Y Acad Sci* 2005;**1043**:777–783.
  107. Stirban A, Negrean M, Stratmann B, et al. Benfotiamine prevents macro- and microvascular endothelial dysfunction and oxidative stress following a meal rich in advanced glycation end products in individuals with type 2 diabetes. *Diabetes Care* 2006;**29**:2064–2071.
  108. Hammes HP, Du X, Edelstein D, et al. Benfotiamine blocks three major pathways of hyperglycemic damage and prevents experimental diabetic retinopathy. *Nat Med* 2003;**9**:294–299.
  109. Polizzi FC, Andican G, Cetin E, Civelek S, Yumuk V, Burçak G. Increased DNA-Glycation in type 2 diabetic patients: the effect of thiamine and pyridoxine therapy. *Exp Clin Endocrinol Diabetes* 2012;**120**:329–334.
  110. Hunot S, Brugg B, Ricard D, et al. Nuclear translocation of NF-kappaB is increased in dopaminergic neurons of patients with parkinson disease. *Proc Natl Acad Sci USA* 1997;**94**:7531–7536.
  111. Soós J, Engelhardt JJ, Siklós L, Havas L, Majtényi K. The expression of PARP, NF-kappa B and parvalbumin is increased in Parkinson disease. *NeuroReport* 2004;**15**:1715–1718.
  112. Togo T, Iseki E, Marui W, Akiyama H, Ueda K, Kosaka K. Glial involvement in the degeneration process of Lewy body-bearing neurons and the degradation process of Lewy bodies in brains of dementia with Lewy bodies. *J Neurol Sci* 2001;**184**:71–75.
  113. Aoki E, Yano R, Yokoyama H, Kato H, Araki T. Role of nuclear transcription factor kappa B (NF-kappaB) for MPTP (1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine)-induced apoptosis in nigral neurons of mice. *Exp Mol Pathol* 2009;**86**:57–64.
  114. Prabhakaran K, Chapman GD, Gunasekar PG.  $\alpha$ -Synuclein overexpression enhances manganese-induced neurotoxicity through the NF- $\kappa$ B-mediated pathway. *Toxicol Mech Methods* 2011;**21**:435–443.
  115. Ghosh A, Roy A, Liu X, et al. Selective inhibition of NF-kappaB activation prevents dopaminergic neuronal loss in a mouse model of Parkinson's disease. *Proc Natl Acad Sci USA* 2007;**104**:18754–18759.
  116. Yadav UC, Subramanyam S, Ramana KV. Prevention of endotoxin-induced uveitis in rats by benfotiamine, a lipophilic analogue of vitamin B1. *Invest Ophthalmol Vis Sci* 2009;**50**:2276–2282.
  117. Shoen M, Ramana KV. Anti-inflammatory effects of benfotiamine are mediated through the regulation of the arachidonic acid pathway in macrophages. *Free Radic Biol Med* 2012;**52**:182–190.
  118. Hipskind RA, Bilbe G. MAP kinase signaling cascades and gene expression in osteoblasts. *Front Biosci* 1998;**3**:d804–d816.
  119. Zhu JH, Kulich SM, Oury TD, Chu CT. Cytoplasmic aggregates of phosphorylated extracellular signal-regulated protein kinases in Lewy body diseases. *Am J Pathol* 2002;**161**:2087–2098.
  120. Zhu JH, Guo F, Shelburne J, Watkins S, Chu CT. Localization of phosphorylated ERK/MAP kinases to mitochondria and autophagosomes in Lewy body diseases. *Brain Pathol* 2003;**13**:473–481.
  121. Kulich SM, Chu CT. Sustained extracellular signal-regulated kinase activation by 6-hydroxydopamine: implications for Parkinson's disease. *J Neurochem* 2001;**77**:1058–1066.
  122. Kulich SM, Horbinski C, Patel M, Chu CT. 6-Hydroxydopamine induces mitochondrial ERK activation. *Free Radic Biol Med* 2007;**43**:372–383.
  123. Dagda RK, Zhu J, Kulich SM, Chu CT. Mitochondrially localized ERK2 regulates mitophagy and autophagic cell stress: implications for Parkinson's disease. *Autophagy* 2008;**4**:770–782.
  124. Lynch-Day MA, Mao K, Wang K, Zhao M, Klionsky DJ. The role of autophagy in Parkinson's disease. *Cold Spring Harb Perspect Med* 2012;**2**:a009357.
  125. Karunakaran S, Ravindranath V. Activation of p38 MAPK in the substantia nigra leads to nuclear translocation of NF-kappaB in MPTP-treated mice: implication in Parkinson's disease. *J Neurochem* 2009;**109**:17911–17919.
  126. Izumi Y, Yamamoto N, Matsuo T, et al. Vulnerability to glutamate toxicity of dopaminergic neurons is dependent on endogenous dopamine and MAPK activation. *J Neurochem* 2009;**110**:745–755.
  127. Thomas T, Timmer M, Cesnulevicius K, Hitti E, Kotlyarov A, Gaestel M. MAPKAP kinase 2-deficiency prevents neurons from cell death by reducing neuroinflammation—relevance in a mouse model of Parkinson's disease. *J Neurochem* 2008;**105**:2039–2052.
  128. Choi DH, Cristóvão AC, Guhathakurta S, et al. NADPH oxidase 1-mediated oxidative stress leads to dopamine neuron death in Parkinson's disease. *Antioxid Redox Signal* 2012;**16**:1033–1045.
  129. Kumar A, Singh BK, Ahmad I, et al. Involvement of NADPH oxidase and glutathione in zinc-induced dopaminergic neurodegeneration in rats: similarity with paraquat neurotoxicity. *Brain Res* 2012;**1438**:48–64.
  130. Zawada WM, Banninger GP, Thornton J, et al. Generation of reactive oxygen species in 1-methyl-4-phenylpyridinium (MPP+) treated dopaminergic neurons occurs as an NADPH oxidase-dependent two-wave cascade. *J Neuroinflammation* 2011;**8**:129.
  131. Zhou H, Zhang F, Chen SH, et al. Rotenone activates phagocyte NADPH oxidase by binding to its membrane subunit gp91phox. *Free Radic Biol Med* 2012;**52**:303–313.
  132. Rodríguez-Pallares J, Rey P, Parga JA, Muñoz A, Guerra MJ, Labandeira-García JL. Brain angiotensin enhances dopaminergic cell death via microglial activation and NADPH-derived ROS. *Neurobiol Dis* 2008;**31**:58–73.
  133. Cristóvão AC, Choi DH, Baltazar G, Beal MF, Kim YS. The role of NADPH oxidase 1-derived reactive oxygen

- species in paraquat-mediated dopaminergic cell death. *Antioxid Redox Signal* 2009;**11**:2105–2118.
134. Anantharam V, Kaul S, Song C, Kanthasamy A, Kanthasamy AG. Pharmacological inhibition of neuronal NADPH oxidase protects against 1-methyl-4-phenylpyridinium (MPP+)-induced oxidative stress and apoptosis in mesencephalic dopaminergic neuronal cells. *Neurotoxicology* 2007;**28**:988–997.
  135. Zhang D, Hu X, Wei SJ, et al. Squamosamide derivative FLZ protects dopaminergic neurons against inflammation-mediated neurodegeneration through the inhibition of NADPH oxidase activity. *J Neuroinflammation* 2008;**5**:21.
  136. Shangari N, Mehta R, O'Brien PJ. Hepatocyte susceptibility to glyoxal is dependent on cell thiamin content. *Chem Biol Interact* 2007;**165**:146–154.
  137. Galdhar NR, Pawar SS. Hepatic drug metabolism and lipid peroxidation in thiamine deficient rats. *Int J Vitam Nutr Res* 1976;**46**:14–23.
  138. Fraser DA, Hessvik NP, Nikolić N, et al. Benfotiamine increases glucose oxidation and downregulates NADPH oxidase 4 expression in cultured human myotubes exposed to both normal and high glucose concentrations. *Genes Nutr* 2012;**7**:459–469.
  139. Grosse W 3rd, Wade AE. The effect of thiamine consumption on liver microsomal drug-metabolizing pathways. *J Pharmacol Exp Ther* 1971;**176**:758–765.
  140. Pfeiffer RF. Gastrointestinal dysfunction in Parkinson's disease. *Lancet Neurol* 2003;**2**:107–116.
  141. Heetun ZS, Quigley EM. Gastroparesis and Parkinson's disease: a systematic review. *Parkinsonism Relat Disord* 2012;**18**:433–440.
  142. Davies KN, King D, Billington D, Barrett JA. Intestinal permeability and orocaecal transit time in elderly patients with Parkinson's disease. *Postgrad Med J* 1996;**72**:164–167.
  143. Baum RA, Iber FL. Thiamin—the interaction of aging, alcoholism, and malabsorption in various populations. *World Rev Nutr Diet* 1984;**44**:85–116.